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(71) Applicant:
**Max-Planck-Gesellschaft zur Förderung
der Wissenschaften e.V. Berlin
80539 München (DE)**

(72) Inventors:
• **Meyer, Thomas, Prof.Dr.
72076 Tübingen (DE)**

• **Gómez, Oscar, Dr.
72076 Tübingen (DE)**
• **Yan, Zhengxin, Dr.
72076 Tübingen (DE)**
• **Haas, Rainer, Dr.
72076 Tübingen (DE)**
• **Lucas, Bernadette, Dr.
72076 Tübingen (DE)**

(74) Representative:
**Weiss, Wolfgang, Dipl.-Chem. Dr. et al
Patentanwälte
Weickmann & Partner,
Kopernikusstrasse 9
81679 München (DE)**

(54) **Helicobacter pylori live vaccine**

(57) The present invention relates to novel recombinant live vaccines, which provide protective immunity against an infection by *Helicobacter pylori* and a method of screening *H. pylori* antigens for optimized vaccines.

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Description

The present invention relates to novel recombinant live vaccines, which provide protective immunity against an infection by *Helicobacter pylori* and a method of screening *H. pylori* antigens for optimized vaccines.

5 *Helicobacter* is a gram-negative bacterial pathogen associated with the development of gastritis, peptic ulceration and gastric carcinoma. Several *Helicobacter* species colonize the stomach, most notably *H. pylori*, *H. heilmanii* and *H. felis*. Although *H. pylori* is the species most commonly associated with human infection, *H. heilmanii* and *H. felis* also have been found to infect humans. High *H. pylori* infection rates are observed in third world countries, as well as in industrialized countries. Among all the virulence factors described in *H. pylori*, urease is known to be essential for colonisation of gnotobiotic pigs and nude mice. Urease is an enzyme composed of two structural subunits (UreA and UreB).
10 Previous studies have indicated that oral immunization using recombinant UreB plus cholera toxin were able to protect mice from gastric colonisation with *H. felis* and *H. pylori* (Michetti et al., *Gastroenterology* 107 (1994), 1002-1011). By oral administration of recombinant UreB antigens, however, in several cases only an incomplete protection can be obtained. Other *H. pylori* antigens shown to give partial protection are the 87 kD vacuolar cytotoxin VacA (Cover and Blaser, *J. Biol. Chem.* 267 (1992), 10570; Marchetti et al., *Science* 267 (1995), 1655) and the 13 and 58 kD heat shock proteins HspA and HspB (Ferrero et al., *Proc. Natl. Acad. Sci. USA* 92 (1995), 6499).

Attenuated pathogens, e.g. bacteria, such as *Salmonella*, are known to be efficient live vaccines. The first indications of the efficacy of attenuated *Salmonella* as good vaccine in humans came from studies using a chemically mutagenized *Salmonella typhi* Ty21a strain (Germanier and Furer, *J. Infect. Dis.* 141 (1975), 553-558), tested successfully
20 in adult volunteers (Gilman et al., *J. Infect. Dis.* 136 (1977), 717-723) and later on in children in a large field trial in Egypt (Whadan et al., *J. Infect. Dis.* 145 (1982), 292-295). The orally administered Ty21a vaccine was able to protect 96% of the Egyptian children vaccinated during three years of surveillance. Since that time new attenuated *Salmonella* live vector vaccines have developed (Hone et al., *Vaccine* 9 (1991), 810-816), in which well defined mutations incorporated into the chromosome gave rise to non-virulent strains able to induce strong immune responses after oral administration
25 (Tacket et al., *Vaccine* 10 (1992), 443-446 and Tacket et al., *Infect. Immun.* 60 (1992), 536-541). Other advantages of the live attenuated *Salmonella* vaccine include its safety, easy administration, long-time protection and no adverse reactions in comparison with the former inactivated wholesale typhoid vaccines (Levine et al., *Typhoid Fever Vaccines*. In: Plotkin S.A., Mortimer E.A. Jr. (eds.) *Vaccines*. Philadelphia: WB Saunders (1988), 333-361).

Mutants of *S. typhimurium* have been extensively used to deliver antigens because of the possibility to use mice as
30 an animal model, which is believed to mimic *S. typhi* infections in humans. The attenuation of *S. typhimurium* most commonly used consists in site directed mutagenesis of genes affecting the synthesis of aromatic amino acids. Such strains, designated aro mutants, have a negligible pathogenicity, as demonstrated in animal models and human trials using these constructs (Hoiseth and Stocker, *Nature* 291 (1981), 238-239; Tacket et al. (1992), *Supra*). Advantage has been taken from the potent immunogenicity of live *Salmonella* vaccine to deliver heterologous antigens. Expression of
35 specific antigens in attenuated *Salmonella* has conferred murine protection against several bacterial pathogens. The use of recombinant live vaccines, which are capable of expressing *Helicobacter* antigens and protecting the vaccinated animals, has not yet been described.

The use of attenuated live vaccines for the treatment of a *Helicobacter* infection has also not been rendered obvious. The reason therefor being that in the course of the *Helicobacter* infection a strong immune response against the
40 pathogen per se is induced, which, however, does not lead to a protective immunity. Thus, it was highly surprising that a protective immune response is achieved when using recombinant attenuated bacterial cells as antigen carriers, which are capable of expressing a DNA molecule encoding a *Helicobacter* antigen. Apparently, recombinant attenuated bacterial cells expressing a *Helicobacter* antigen are capable of creating a qualitatively different immune response against the heterologous *Helicobacter* antigen than *Helicobacter* itself does against its own homologous antigen. Surprisingly,
45 a non-protective immune response is thus transformed into an immune response protecting against *Helicobacter* infections. This unexpected observation renders it possible to use recombinant attenuated pathogens, e.g. bacterial cells, particularly *Salmonella*, as carriers for the screening of protective antigens, to apply the protective antigens identified in this manner in any vaccine against *Helicobacter* infections, and to use recombinant attenuated bacteria as carriers of protective antigens for the immunization against *Helicobacter* infections in humans and other mammals.

50 Thus, a subject matter of the present invention is a recombinant attenuated pathogen, which comprises at least one heterologous nucleic acid molecule encoding a *Helicobacter* antigen, wherein said pathogen is capable to express said nucleic acid molecule or capable to cause the expression of said nucleic acid in a target cell. Preferably the nucleic acid molecule is a DNA molecule.

The attenuated pathogen is a microorganism strain which is able to cause infection and preferably effective immunological protection against the actual pathogen but is no longer pathogenic per se. The attenuated pathogen can be a
55 bacterium, a virus, a fungus or a parasite. Preferably it is a bacterium, e.g. *Salmonella*, such as *S. typhimurium* or *S. typhi*, *Vibrio cholerae* (Mekalanos et al., *Nature* 306 (1983), 551-557), *Shigella* Species such as *S. flexneri* (Sizemore et al., *Science* 270 (1995), 299-302; Mounier et al., *EMBO J.* 11 (1992), 1991-1999), *Listeria* such as *L. monocytogenes*

(Milon and Cossart, Trends in Microbiology 3 (1995), 451-453), Escherichia coli, Streptococcus, such as S. gordonii (Medaglini et al., Proc. Natl. Acad. Sci. USA 92 (1995) 6868-6872) or Mycobacterium, such as Bacille Calmette Guérin (Flynn, Cell. Mol. Biol. 40 Suppl. 1 (1994), 31-36). More preferably the pathogen is an attenuated enterobacterium such as Vibrio cholerae, Shigella flexneri, Escherichia coli or Salmonella. Most preferably the attenuated pathogen is a Salmonella cell, e.g. a Salmonella aro mutant cell. The attenuated pathogen, however, can be a virus, e.g. an attenuated vaccinia virus, adenovirus or pox virus.

The nucleic acid molecule which is inserted into the pathogen codes for a Helicobacter antigen, preferably a H. felis, H. heilmanii or H. pylori antigen, more preferably a H. pylori antigen. The Helicobacter antigen can be a native Helicobacter polypeptide, an immunologically reactive fragment thereof, or an immunologically reactive variant of a native polypeptide or of a fragment thereof. Further, the Helicobacter antigen can be a protective carbohydrate or a peptide mimotope simulating the three-dimensional structure of a native Helicobacter antigen. Peptide mimotopes can be obtained from peptide libraries presented on the surface of bacterial cells (cf. PCT/EP96/01130). Of course, the transformed cell can also contain several DNA molecules coding for different Helicobacter antigens.

Attenuated bacteria can be used to transcribe and translate said nucleic acid molecule directly in the bacterial cell or to deliver said nucleic acid molecule to the infected target cell, such that the DNA molecule is transcribed and/or translated by the eukaryotic target cell machinery. This indirect bacterial vaccination procedure, termed here as genetic vaccination, has been successfully used with Shigella as a carrier (Sizemore, D. R., Branstrom, A. A. & Sadoff, J. C. (1995) Attenuated Shigella as a DNA delivery vehicle for DNA-mediated immunization. Science 270:299-302).

In a preferred embodiment of the present invention the Helicobacter antigen is urease, a urease subunit or an immunologically reactive variant or fragment thereof or a peptide mimotope thereof. In a further preferred embodiment of the present invention the Helicobacter antigen is a secretory polypeptide from Helicobacter, an immunologically reactive variant or fragment thereof or a peptide mimotope thereof. A process for identifying Helicobacter genes coding for such secretory polypeptides, and particularly for adhesins, has been disclosed in the international patent application PCT/EP96/02544, which is incorporated herein by reference. This process comprises

- a) preparing a gene bank of H. pylori DNA in a host organism containing an inducible transposon coupled to a marker of secretory activity,
- b) inducing the insertion of the transposon into the H. pylori DNA and
- c) conducting a selection for clones containing a secretory gene by means of the marker, and optionally further
- d) conducting a retransformation of H. pylori by means of the DNA of clones containing genes having secretory activity, wherein isogenic H. pylori mutant strains are produced by means of integrating the DNA into the chromosome, and
- e) conducting a selection detecting adherence-deficient H. pylori mutant strains.

Suitable examples of antigens obtainable by the above process are selected from the group consisting of the antigens AlpA, AlpB, immunologically reactive variants or fragments thereof or peptide mimotopes thereof. The nucleic and amino acid sequences of the antigens AlpA and AlpB have been disclosed in the international patent applications PCT/EP96/02545 and PCT/EP96/04124, which are incorporated herein by reference. Further, the nucleic and amino acid sequences of AlpB are shown in SEQ ID NO. 1 and 2, and the nucleic and amino acid sequences of AlpA in SEQ ID NO. 3 and 4.

It is also conceivable, however, that an intracellular antigen is used which can be presented on the surface, e.g. by autolytic release, and confers immunological protection.

The presentation of the Helicobacter antigens in the recombinant pathogen according to the invention can be accomplished in different ways. The antigen or the antigens can be synthesized in a constitutive, inducible or phase variable manner in the recombinant pathogen. Concerning the constitutive or inducible synthesis of the Helicobacter antigens known expression systems can be referred to, as have been described by Sambrook et al., Molecular Cloning, A Laboratory Manual (1989), Cold Spring Harbor Laboratory Press.

Particularly preferred the antigens are presented in a phase variable expression system. Such a phase variable expression system for the production and presentation of foreign antigens in hybrid live vaccines is disclosed in EP-B-0 565 548, which is herein incorporated by reference. In such a phase variable expression system the nucleic acid molecule encoding the Helicobacter antigen is under control of an expression signal, which is substantially inactive in the pathogen, and which is capable of being activated by a spontaneous reorganization caused by a nucleic acid, e.g. DNA reorganization mechanism in the pathogen, e.g. a specific DNA inversion process, a specific DNA deletion process, a specific DNA replication process or a specific slipped-strand-mispairing mechanism.

A recombinant cell having a phase variable expression system is capable of forming two subpopulations A and B, wherein the division into said subpopulations occurs by spontaneous reorganization in the recombinant nucleic acid, wherein said sub-population A is capable of infection and immunologically active per se, while subpopulation B, which is regenerated from subpopulation A, produces at least one heterologous Helicobacter antigen and acts immunologi-

cally with respect to said additional antigen.

The activation of the expression signal encoding the Helicobacter antigen can be directly accomplished by nucleic acid reorganization or, alternatively, indirectly accomplished by activation of a gene encoding a protein which controls the expression of the gene encoding the Helicobacter antigen. The indirect activation represents a system which allows the production of the Helicobacter antigen via a cascade system, which can be realized e.g. in that the gene directly controlled by DNA reorganization codes for an RNA polymerase which is specific for the promoter preceding the Helicobacter gene, or a gene regulator which in another specific manner induces the expression of the Helicobacter gene. In an especially preferred embodiment of the present invention the expression signal for the gene encoding the Helicobacter antigen is a bacteriophage promoter, e.g. a T3, T7 or SP6 promoter, and the activation of the expression signal is caused by a nucleic acid reorganization resulting in the production of a corresponding bacteriophage RNA polymerase in the pathogen.

The phase variable expression system can be adjusted to provide a preselected expression level of the Helicobacter antigen. This can be accomplished e.g. by modifying the nucleotide sequence of the expression signal, which is activated by the nucleic acid reorganization mechanism, and/or by inserting further genetic regulation elements.

The Helicobacter antigens can be produced in an intracellular, as well as in an extracellular manner in the pathogen according to the invention. For instance, autotransporter systems such as the IgA-protease system (cf. for instance EP-A-0 254 090) or the E. coli AIDA-1 adhesin system (Benz et al., Mol. Microbiol. 6 (1992), 1539) are suited as extracellular secretory system. Other suitable outer membrane transporter systems are the RTX-toxin transporters, e.g. the E. coli hemolysin transport system (Hess et al., Proc. Natl. Acad. Sci. USA 93 (1996), 11458-11463).

The pathogen according to the invention can contain a second heterologous nucleic acid, e.g. DNA molecule, which codes for an immunomodulatory polypeptide influencing the immune response quantitatively or qualitatively, apart from the nucleic acid molecule encoding the Helicobacter antigen. Examples of such immunomodulatory polypeptides are immune-stimulating peptides, cytokines like IL-2, IL-6 or IL-12, chemokines, toxins, such as cholera toxin B or adhesins.

The present invention also refers to a pharmaceutical composition comprising as an active agent a recombinant attenuated pathogen as described above, optionally together with pharmaceutically acceptable diluents, carriers and adjuvants. Preferably, the composition is a living vaccine. The vaccination routes depend upon the choice of the vaccination vector. The administration may be achieved in a single dose or repeated at intervals. The appropriate dosage depends on various parameters such as the vaccinal vector itself, or the route of administration. Administration to a mucosal surface (e.g. ocular, intranasal, oral, gastric, intestinal, rectal, vaginal or urinary tract) or via the parenteral route (e.g. subcutaneous, intradermal, intramuscular, intravenous or intraperitoneal) might be chosen. A method for the preparation of the living vaccine comprises formulating the attenuated pathogen in a pharmaceutically effective amount with pharmaceutically acceptable diluents, carriers and/or adjuvants.

Further, the present invention refers to a method for preparing a recombinant attenuated pathogen as defined above, comprising the steps of a) inserting a nucleic acid molecule encoding a Helicobacter antigen into an attenuated pathogen, wherein the recombinant pathogen, e.g. a transformed bacterial cell, is obtained, which is capable of expressing said nucleic acid molecule or is capable to cause expression of said nucleic acid molecule in a target cell and b) cultivating said recombinant attenuated pathogen under suitable conditions. If the pathogen is a bacterial cell, the nucleic acid molecule encoding the Helicobacter antigen can be located on an extrachromosomal plasmid. It is, however, also possible to insert the nucleic acid molecule into the chromosome of the pathogen.

Furthermore, the present invention refers to a method for identifying Helicobacter antigens which raise a protective immune response in a mammalian host, comprising the steps of:

a) providing an expression gene bank of Helicobacter in an attenuated pathogen and b) screening the clones of the gene bank for the ability to confer a protective immunity against a Helicobacter infection in a mammalian host. Preferably, this identification process takes place in a phase variable expression system, rendering possible a stable expression of all of the Helicobacter antigens. Recombinant clones can then be applied as "pools" for the oral immunization of test animals, such as mice. The potential of these clones as protective antigen is then determined via a challenge infection with Helicobacter, e.g. a mouse-adapted H. pylori strain. Thus, there is a possibility of directly selecting optimized H. pylori vaccine antigens.

The invention will be further illustrated by the following figures and sequence listings.

Fig. 1: shows a schematic illustration of the urease expression vector pYZ97, whereon the genes coding for the urease subunits UreA and UreB are located under transcriptional control of the T7 promoter $\phi 10$. There is a ribosomal binding site (RBS) between the T7 promoter and the urease genes. Further, the plasmid exhibits an origin of replication (ori), a β -lactamase resistance gene (bla) and 4 T7 terminators in series. Apart from the expression by the T7 promoter, a constitutive low level expression of the urease A and B sub-

units can also be brought about via a cryptic promoter, which is located upstream from the T7 promoter, on the plasmid pYZ97.

Fig. 2: shows a schematic illustration of the T7 RNA polymerase (T7RNAP) expression cassettes pYZ88, pYZ84 and pYZ114, which can be integrated into the chromosomes of bacteria.

In the high-expression cassette pYZ88 the lambda PL promoter is located in inverse orientation, upstream from the T7RNAP gene. A gene for the temperature-sensitive repressor *ci 857* (*ci*) is under control of this promoter. A terminator of the bacteriophage *fd* (*fdT*) is situated upstream from the *ci* gene. The *gin* gene (Mertens, EMBO J. 3 (1984), 2415-2421) codes for a control enzyme of a DNA reorganization mechanism. A DNA sequence coding for the tRNA Arg is located downstream from the *gin* gene.

In phase A the PL promoter responsible for the expression of the T7RNAP gene is directed in the direction of the *ci857* gene and the *gin* gene. The consequence of this is that an active repressor is formed at the permissive temperature of 28°C and reduces the transcription from the PL promoter. At a higher temperature the transcription of the PL promoter is increased, since the repressor is inactivated at least partially under such external influences. The temperature-dependent increase in the transcription also causes a corresponding increase in the expression of the following *gin* gene, which as a control enzyme catalyses the inversion of the PL promoter and the transition in phase B, in which the T7RNAP gene is expressed.

In the high-expression system pYZ88 a further *fdT* transcription terminator is located between a kanamycin-resistance gene (*km*) and the promoter of this gene. In this manner, the synthesis of an anti-sense RNA, inversely orientated to the T7RNAP gene, which normally contributes to the reduction of the T7RNAP expression, is reduced. This results in a high expression of the T7RNAP.

In the medium-expression system pYZ84 a transcription terminator (*fdT*) is located between the PL promoter and the start of the T7RNAP gene. In this manner the expression of the T7RNAP mRNA is reduced. Additionally, the anti-sense RNA affects the T7RNAP translation. Therefore, only a medium expression occurs.

In the low-expression system pYZ114 a deletion of 100 bp in PL is additionally introduced (Δ PL). In this manner the activity of the PL promoter is reduced to a high extent, which leads to a lower T7RNAP expression and thus to a reduction of the *UreA/B* gene expression. In this construct the effect of the cryptic promoter on pYZ97 is already observed.

SEQ ID NO. 1 and 2 show the nucleotide sequence of the adhesin gene *AlpB* from *H. pylori* and the amino acid sequence of the polypeptide coded therefrom.

SEQ ID NO. 3 and 4 show the nucleotide sequence of the adhesin gene *AlpA* from *H. pylori* and the amino acid sequence of the protein coded therefrom.

Experimental part

Materials and Methods

Bacterial strains: *S. typhimurium* SL3261 live vector vaccine strain was used as a recipient for the recombinant *H. pylori* urease plasmid constructs. *S. typhimurium* SL3261 is an *aroA* transposon mutant derived from *S. typhimurium* SL1344 wild type strain. *S. typhimurium* SL3261 is a non-virulent strain that gives protection to mice against infection with wild type *S. typhimurium* after oral administration (Hoiseth and Stocker (1981) Supra). *S. typhimurium* SL3261 and derivatives thereof, which contain the urease expression plasmid pYZ97 (extrachromosomal) and the T7RNAP expression cassettes pYZ88, pYZ84 or pYZ114, respectively (integrated into the chromosome) are indicated in table 1. Luria broth or agar was used for bacterial growth at 28°C. *H. pylori* wild type strain grown at 37°C on serum plates was used for the challenge experiments.

Immunization of mice: Four weeks Balb/c mice purchased from Interfauna (Tuttlingen, Germany) were adapted two weeks in an animal facility before being used for experimentation. 150 μ l of blood was taken retroorbitally from all mice to obtain preimmune serum. Retroorbital bleedings were repeated from all immunized mice 1 week and 3 weeks after immunization.

Eight groups of 5 mice including controls were used in this study (table 2). Group A, the naive control group, was not immunized with *Salmonella* neither challenged with wild type *H. pylori*. The rest of the groups were all orally immunized. Group B, a negative control group, did not receive *Salmonella* and was challenged with *H. pylori*. Mice from groups C to G were immunized with *Salmonella* vaccine strains and challenged with *H. pylori*. The last group H received recombinant urease B in combination with cholera toxin and was also challenged.

Prior to immunizations mice were left overnight without solid food and 4 hours without water. 100 μ l of 3% sodium

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bicarbonate were given orally using a stainless steel catheter tube to neutralize the stomach pH. Then mice from group B received 100 µl PBS and mice from groups C to G received 1.0×10^{10} CFU of *Salmonella* in a 100 µl volume. Mice from group H received four times 100 µl of a mixture of recombinant *H. pylori* UreaseB plus cholera toxin, one dose every week. After every immunization water and food were returned to the mice.

H. pylori challenge: Four weeks after the first oral immunization mice from groups B to H were challenged with *H. pylori*. Mice were left overnight without solid food and without water 4 hours prior to the challenge. 100 µl of 3% sodium bicarbonate were given orally to the mice using a stainless steel catheter tube, followed by an oral dose of 5.0×10^9 CFU/ml of *Helicobacter pylori*. Water and food were returned to the mice after the challenge.

Collection of blood and tissues from mice: Twelve weeks after the first immunization the mice were left overnight without food and subsequently sacrificed for analysis of protection and immune response. The mice were anaesthetized with Metoxyfluorane for terminal cardiac bleeding and prior to sacrifice by cervical dislocation. Under aseptic conditions, spleen and stomach were carefully removed from each mouse and placed on ice in separate sterile containers until further processing. Large and small intestine were obtained for further isolation of the intestinal fluid.

Processing of stomach and measurement of urease activity: The degree of *H. pylori* colonisation in the mouse stomach was measured by the presence of active urease in the tissue. The Jatrox-test (Röhm-Pharma GmbH, Weiterstadt, Germany) was used according to the suppliers' directions. Stomach mucosa was exposed and washed with PBS, half of the antral portion of the stomach was immediately placed inside an Eppendorf tube containing the substrate for measurement of urease activity. Absorbance at 550 nm was measured after tubes were incubated for 4 hours at room temperature. The rest of the stomach tissue was stored at -20°C for further treatments. The urease activity values obtained from the stomach of naive mice, which did not undergo immunization or challenge, were used to create a base line to indicate the absence of *H. pylori* infection and therefore protection.

Table 1

UreA and UreB expressing <i>S. typhimurium</i> vaccine strains		
Strains	Urease Expression	Source
<i>S. typhimurium</i> SL3261	Negative	Hoiseth and Stocker
<i>S. typhimurium</i> SL3262 pYZ97	Constitutive Low	this study
<i>S. typhimurium</i> SL3261::pYZ88pYZ97	High T7-induced expression	this study
<i>S. typhimurium</i> SL3261::pYZ84pYZ97	Medium T7-induced expression	this study
<i>S. typhimurium</i> SL3261::pYZ114pYZ97	Low T7-induced expression	this study

Table 2

Mice groups used for immunization		
Group	Immunogen	No. of oral immunizations
A	None	0
B	PBS oral immunization	1
C	<i>S. typhimurium</i> S3261	1
D	<i>S. typhimurium</i> S3261 pYZ97	1
E	<i>S. typhimurium</i> S3261::pYZ88pYZ97	1
F	<i>S. typhimurium</i> S3261::pYZ84pYZ97	1
G	<i>S. typhimurium</i> S3261::pYZ114pYZ97	1
H	Urease B plus cholera toxin	4

Results:

In the control mice (groups B and C) 100% infection with *H. pylori* was observed. In the mice immunized with recombinant attenuated pathogens (groups D, E, F, G) between 0% and 60% infection (100% to 40% protection) was observed. Immuno-protection did not correlate with humoral anti-UreA and UreB response, suggesting that, in addition to humoral immunity, cellular immunity is critical for protection against *H. pylori* infection. The results indicate that oral immunization of mice with UreA and UreB delivered by *S. typhimurium* attenuated strain is effective to induce high levels of protection against *H. pylori* colonisation.

In the mice immunized with recombinant urease B plus cholera toxin considerably higher levels of urease activity were observed under said experimental conditions than when administering the recombinant attenuated pathogens according to the invention.

The results of the urease test have been illustrated in table 3.

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Table 3

	Group	Mouse	E _{550nm, 4h}	E _{4h} - E _{control}	E _{ser.} * 3	Dilution
5	A	1	0,085	-0,022	-0,066	200µl+400µl
	A	2	0,091	-0,016	-0,048	200µl+400µl
	A	3	0,116	0,009	0,027	200µl+400µl
	A	4	0,099	-0,008	-0,024	200µl+400µl
	A	5	0,101	-0,006	-0,018	200µl+400µl
	Control		0,107	0	0	200µl+400µl
10	B	1	0,394	0,292	0,876	200µl+400µl
	B	2	0,464	0,362	1,086	200µl+400µl
	B	3	0,329	0,227	0,681	200µl+400µl
	B	4	0,527	0,425	1,275	200µl+400µl
	B	5	0,462	0,36	1,08	200µl+400µl
	Control		0,102	0	0	200µl+400µl
15	C	1	0,248	0,145	0,435	200µl+400µl
	C	2	0,369	0,266	0,798	200µl+400µl
	C	3	0,209	0,106	0,318	200µl+400µl
	C	4	0,219	0,116	0,348	200µl+400µl
	C	5	0,24	0,137	0,411	200µl+400µl
	Control		0,103	0	0	200µl+400µl
20	D	1	0,143	0,002	0,004	300µl+300µl
	D	2	0,156	0,015	0,03	300µl+300µl
	D	3	0,142	0,001	0,002	300µl+300µl
	D	4	0,114	-0,027	-0,054	300µl+300µl
	D	5	0,133	-0,008	-0,016	300µl+300µl
	Control		0,141	0	0	300µl+300µl
25	E	1	0,127	0,027	0,081	200µl+400µl
	E	2	0,094	-0,006	-0,018	200µl+400µl
	E	3	0,099	-0,001	-0,003	200µl+400µl
	E	4	0,161	0,061	0,183	200µl+400µl
	E	5	0,198	0,098	0,294	200µl+400µl
	Control		0,1	0	0	200µl+400µl
30	F	1	0,166	0,025	0,05	300µl+300µl
	F	2	0,145	0,004	0,008	300µl+300µl
	F	3	0,166	0,025	0,05	300µl+300µl
	F	4	0,154	0,013	0,026	300µl+300µl
	F	5	0,301	0,16	0,32	300µl+300µl
	Control		0,141	0	0	300µl+300µl
35	G	1	0,084	-0,019	-0,057	200µl+400µl
	G	2	0,087	-0,016	-0,048	200µl+400µl
	G	3	0,269	0,166	0,498	200µl+400µl
	G	4	0,085	-0,018	-0,054	200µl+400µl
	G	5	0,092	-0,011	-0,033	200µl+400µl
	Control		0,103	0	0	200µl+400µl
40	H	1	0,638	0,531	1,593	200µl+400µl
	H	2	0,282	0,175	0,525	200µl+400µl
	H	3	0,141	0,034	0,102	200µl+400µl
	H	4	0,135	0,028	0,084	200µl+400µl
	H	5	0,171	0,064	0,192	200µl+400µl
	Control		0,107	0	0	200µl+400µl
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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: Max-Planck-Gesellschaft zur Foerderung der
Wissenschaften e.V. Berlin
- (B) STREET: Hofgartenstr. 2
- (C) CITY: Muenchen
- (E) COUNTRY: Germany
- (F) POSTAL CODE (ZIP): 80539

(ii) TITLE OF INVENTION: Helicobacter pylori live vaccine

(iii) NUMBER OF SEQUENCES: 4

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1557 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: both
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Helicobacter pylori

(vii) IMMEDIATE SOURCE:

- (B) CLONE: alpB

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1554

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

40	ATG ACA CAA TCT CAA AAA GTA AGA TTC TTA GCC CCT TTA AGC CTA GCG	48
	Met Thr Gln Ser Gln Lys Val Arg Phe Leu Ala Pro Leu Ser Leu Ala	
	1 5 10 15	
	TTA AGC TTG AGC TTC AAT CCA GTG GGC GCT GAA GAA GAT GGG GGC TTT	96
	Leu Ser Leu Ser Phe Asn Pro Val Gly Ala Glu Glu Asp Gly Gly Phe	
	20 25 30	
45	ATG ACC TTT GGG TAT GAA TTA GGT CAG GTG GTC CAA CAA GTG AAA AAC	144
	Met Thr Phe Gly Tyr Glu Leu Gly Gln Val Val Gln Gln Val Lys Asn	
	35 40 45	
50	CCG GGT AAA ATC AAA GCC GAA GAA TTA GCC GGC TTG TTA AAC TCT ACC	192
	Pro Gly Lys Ile Lys Ala Glu Glu Leu Ala Gly Leu Leu Asn Ser Thr	
	50 55 60	
	ACA ACA AAC AAC ACC AAT ATC AAT ATT GCA GGC ACA GGA GGC AAT GTC	240

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	Thr	Leu	Ala	Lys	Val	Ser	Ala	Leu	Asn	Asn	Glu	Leu	Lys	Ala	Asn	Pro	
					325					330					335		
5	TGG	CTT	GGG	AAT	TTT	GCC	GCC	GGT	AAC	AGC	TCT	CAA	GTG	AAT	GCG	TTT	1056
	Trp	Leu	Gly	Asn	Phe	Ala	Ala	Gly	Asn	Ser	Ser	Gln	Val	Asn	Ala	Phe	
				340					345					350			
	AAC	GGG	TTT	ATC	ACT	AAA	ATC	GGT	TAC	AAG	CAA	TTC	TTT	GGG	GAA	AAC	1104
	Asn	Gly	Phe	Ile	Thr	Lys	Ile	Gly	Tyr	Lys	Gln	Phe	Phe	Gly	Glu	Asn	
				355				360					365				
10	AAG	AAT	GTG	GGC	TTA	CGC	TAC	TAC	GGC	TTC	TTC	AGC	TAT	AAC	GGC	GCG	1152
	Lys	Asn	Val	Gly	Leu	Arg	Tyr	Tyr	Gly	Phe	Phe	Ser	Tyr	Asn	Gly	Ala	
		370					375					380					
15	GGC	GTG	GGT	AAT	GGC	CCT	ACT	TAC	AAT	CAA	GTC	AAT	TTG	CTC	ACT	TAT	1200
	Gly	Val	Gly	Asn	Gly	Pro	Thr	Tyr	Asn	Gln	Val	Asn	Leu	Leu	Thr	Tyr	
		385				390					395				400		
	GGG	GTG	GGG	ACT	GAT	GTG	CTT	TAC	AAT	GTG	TTT	AGC	CGC	TCT	TTT	GGT	1248
	Gly	Val	Gly	Thr	Asp	Val	Leu	Tyr	Asn	Val	Phe	Ser	Arg	Ser	Phe	Gly	
				405						410					415		
20	AGT	AGG	AGT	CTT	AAT	GCG	GGC	TTC	TTT	GGG	GGG	ATC	CAA	CTC	GCA	GGG	1296
	Ser	Arg	Ser	Leu	Asn	Ala	Gly	Phe	Phe	Gly	Gly	Ile	Gln	Leu	Ala	Gly	
				420					425					430			
25	GAT	ACT	TAC	ATC	AGC	ACG	CTA	AGA	AAC	AGC	TCT	CAG	CTT	GCG	AGC	AGA	1344
	Asp	Thr	Tyr	Ile	Ser	Thr	Leu	Arg	Asn	Ser	Ser	Gln	Leu	Ala	Ser	Arg	
			435					440					445				
	CCT	ACA	GCG	ACG	AAA	TTC	CAA	TTC	TTG	TTT	GAT	GTG	GGC	TTA	CGC	ATG	1392
	Pro	Thr	Ala	Thr	Lys	Phe	Gln	Phe	Leu	Phe	Asp	Val	Gly	Leu	Arg	Met	
		450				455						460					
30	AAC	TTT	GGT	ATC	TTG	AAA	AAA	GAC	TTG	AAA	AGC	CAT	AAC	CAG	CAT	TCT	1440
	Asn	Phe	Gly	Ile	Leu	Lys	Lys	Asp	Leu	Lys	Ser	His	Asn	Gln	His	Ser	
		465				470					475				480		
35	ATA	GAA	ATC	GGT	GTG	CAA	ATC	CCT	ACG	ATT	TAC	AAC	ACT	TAC	TAT	AAA	1488
	Ile	Glu	Ile	Gly	Val	Gln	Ile	Pro	Thr	Ile	Tyr	Asn	Thr	Tyr	Tyr	Lys	
				485						490					495		
	GCT	GGC	GGT	GCT	GAA	GTG	AAA	TAC	TTC	CGC	CCT	TAT	AGC	GTG	TAT	TGG	1536
	Ala	Gly	Gly	Ala	Glu	Val	Lys	Tyr	Phe	Arg	Pro	Tyr	Ser	Val	Tyr	Trp	
				500					505					510			
40	GTC	TAT	GGC	TAC	GCC	TTC	TAA										1557
	Val	Tyr	Gly	Tyr	Ala	Phe											
				515													

(2) INFORMATION FOR SEQ ID NO: 2:

- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 518 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Thr Gln Ser Gln Lys Val Arg Phe Leu Ala Pro Leu Ser Leu Ala
1 5 10 15

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Leu Ser Leu Ser Phe Asn Pro Val Gly Ala Glu Glu Asp Gly Gly Phe
 20 25 30
 5 Met Thr Phe Gly Tyr Glu Leu Gly Gln Val Val Gln Gln Val Lys Asn
 35 40 45
 Pro Gly Lys Ile Lys Ala Glu Glu Leu Ala Gly Leu Leu Asn Ser Thr
 50 55 60
 10 Thr Thr Asn Asn Thr Asn Ile Asn Ile Ala Gly Thr Gly Gly Asn Val
 65 70 75 80
 Ala Gly Thr Leu Gly Asn Leu Phe Met Asn Gln Leu Gly Asn Leu Ile
 85 90 95
 15 Asp Leu Tyr Pro Thr Leu Asn Thr Ser Asn Ile Thr Gln Cys Gly Thr
 100 105 110
 Thr Asn Ser Gly Ser Ser Ser Ser Gly Gly Gly Ala Ala Thr Ala Ala
 115 120 125
 20 Ala Thr Thr Ser Asn Lys Pro Cys Phe Gln Gly Asn Leu Asp Leu Tyr
 130 135 140
 Arg Lys Met Val Asp Ser Ile Lys Thr Leu Ser Gln Asn Ile Ser Lys
 145 150 155 160
 25 Asn Ile Phe Gln Gly Asn Asn Asn Thr Thr Ser Gln Asn Leu Ser Asn
 165 170 175
 Gln Leu Ser Glu Leu Asn Thr Ala Ser Val Tyr Leu Thr Tyr Met Asn
 180 185 190
 30 Ser Phe Leu Asn Ala Asn Asn Gln Ala Gly Gly Ile Phe Gln Asn Asn
 195 200 205
 Thr Asn Gln Ala Tyr Gly Asn Gly Val Thr Ala Gln Gln Ile Ala Tyr
 210 215 220
 35 Ile Leu Lys Gln Ala Ser Ile Thr Met Gly Pro Ser Gly Asp Ser Gly
 225 230 235 240
 Ala Ala Ala Ala Phe Leu Asp Ala Ala Leu Ala Gln His Val Phe Asn
 245 250 255
 40 Ser Ala Asn Ala Gly Asn Asp Leu Ser Ala Lys Glu Phe Thr Ser Leu
 260 265 270
 Val Gln Asn Ile Val Asn Asn Ser Gln Asn Ala Leu Thr Leu Ala Asn
 275 280 285
 45 Asn Ala Asn Ile Ser Asn Ser Thr Gly Tyr Gln Val Ser Tyr Gly Gly
 290 295 300
 Asn Ile Asp Gln Ala Arg Ser Thr Gln Leu Leu Asn Asn Thr Thr Asn
 305 310 315 320
 Thr Leu Ala Lys Val Ser Ala Leu Asn Asn Glu Leu Lys Ala Asn Pro
 325 330 335
 50 Trp Leu Gly Asn Phe Ala Ala Gly Asn Ser Ser Gln Val Asn Ala Phe
 340 345 350
 55

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Asn Gly Phe Ile Thr Lys Ile Gly Tyr Lys Gln Phe Phe Gly Glu Asn
355 360 365

5 Lys Asn Val Gly Leu Arg Tyr Tyr Gly Phe Phe Ser Tyr Asn Gly Ala
370 375 380

Gly Val Gly Asn Gly Pro Thr Tyr Asn Gln Val Asn Leu Leu Thr Tyr
385 390 395 400

10 Gly Val Gly Thr Asp Val Leu Tyr Asn Val Phe Ser Arg Ser Phe Gly
405 410 415

Ser Arg Ser Leu Asn Ala Gly Phe Phe Gly Gly Ile Gln Leu Ala Gly
420 425 430

15 Asp Thr Tyr Ile Ser Thr Leu Arg Asn Ser Ser Gln Leu Ala Ser Arg
435 440 445

Pro Thr Ala Thr Lys Phe Gln Phe Leu Phe Asp Val Gly Leu Arg Met
450 455 460

20 ~~Asn~~ Phe Gly Ile Leu Lys Lys Asp Leu Lys Ser His Asn Gln His Ser
465 470 475 480

Ile Glu Ile Gly Val Gln Ile Pro Thr Ile Tyr Asn Thr Tyr Tyr Lys
485 490 495

25 Ala Gly Gly Ala Glu Val Lys Tyr Phe Arg Pro Tyr Ser Val Tyr Trp
500 505 510

30 Wal Tyr Gly Tyr Ala Phe
515

2(2) INFORMATION FOR SEQ ID NO: 3:

- 30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1557 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
(A) ORGANISM: Helicobacter pylori
- 40 (vii) IMMEDIATE SOURCE:
(B) CLONE: alpA
- (ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1..1554
- 45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATG ATA AAA AAG AAT AGA ACG CTG TTT CTT AGT CTA GCC CTT TGC GCT 48
Met Ile Lys Lys Asn Arg Thr Leu Phe Leu Ser Leu Ala Leu Cys Ala
520 525 530

50 AGC ATA AGT TAT GCC GAA GAT GAT GGA GGG TTT TTC ACC GTC GGT TAT 96
Ser Ile Ser Tyr Ala Glu Asp Asp Gly Gly Phe Phe Thr Val Gly Tyr
535 540 545 550

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	CAG	CTC	GGG	CAA	GTC	ATG	CAA	GAT	GTC	CAA	Asn	CCA	GGC	GGC	GCT	AAA	144
	Gln	Leu	Gly	Gln	Val	Met	Gln	Asp	Val	Gln	Asn	Pro	Gly	Gly	Ala	Lys	
					555						560				565		
5	AGC	GAC	GAA	CTC	GCC	AGA	GAG	CTT	AAC	GCT	GAT	GTA	ACG	AAC	AAC	ATT	192
	Ser	Asp	Glu	Leu	Ala	Arg	Glu	Leu	Asn	Ala	Asp	Val	Thr	Asn	Asn	Ile	
				570					575					580			
10	TTA	AAC	AAC	AAC	ACC	GGA	GGC	AAC	ATC	GCA	GGG	GCG	TTG	AGT	AAC	GCT	240
	Leu	Asn	Asn	Asn	Thr	Gly	Gly	Asn	Ile	Ala	Gly	Ala	Leu	Ser	Asn	Ala	
				585				590					595				
	TTC	TCC	CAA	TAC	CTT	TAT	TCG	CTT	TTA	GGG	GCT	TAC	CCC	ACA	AAA	CTC	288
	Phe	Ser	Gln	Tyr	Leu	Tyr	Ser	Leu	Leu	Gly	Ala	Tyr	Pro	Thr	Lys	Leu	
		600					605					610					
15	AAT	GGT	AGC	GAT	GTG	TCT	GCG	AAC	GCT	CTT	TTA	AGT	GGT	GCG	GTA	GGC	336
	Asn	Gly	Ser	Asp	Val	Ser	Ala	Asn	Ala	Leu	Leu	Ser	Gly	Ala	Val	Gly	
		615				620				625						630	
	TCT	GGG	ACT	TGT	GCG	GCT	GCA	GGG	ACG	GCT	GGT	GGC	ACT	TCT	CTT	AAC	384
20	Ser	Gly	Thr	Cys	Ala	Ala	Ala	Gly	Thr	Ala	Gly	Gly	Thr	Ser	Leu	Asn	
					635					640					645		
	ACT	CAA	AGC	ACT	TGC	ACC	GTT	GCG	GGC	TAT	TAC	TGG	CTC	CCT	AGC	TTG	432
	Thr	Gln	Ser	Thr	Cys	Thr	Val	Ala	Gly	Tyr	Tyr	Trp	Leu	Pro	Ser	Leu	
				650				655						660			
25	ACT	GAC	AGG	ATT	TTA	AGC	ACG	ATC	GGC	AGC	CAG	ACT	AAC	TAC	GGC	ACG	480
	Thr	Asp	Arg	Ile	Leu	Ser	Thr	Ile	Gly	Ser	Gln	Thr	Asn	Tyr	Gly	Thr	
			665					670					675				
	AAC	ACC	AAT	TTC	CCC	AAC	ATG	CAA	CAA	CAG	CTC	ACC	TAC	TTG	AAT	GCG	528
30	Asn	Thr	Asn	Phe	Pro	Asn	Met	Gln	Gln	Gln	Leu	Thr	Tyr	Leu	Asn	Ala	
			680				685					690					
	GGG	AAT	GTG	TTT	TTT	AAT	GCG	ATG	AAT	AAG	GCT	TTA	GAG	AAT	AAG	AAT	576
	Gly	Asn	Val	Phe	Phe	Asn	Ala	Met	Asn	Lys	Ala	Leu	Glu	Asn	Lys	Asn	
						700					705					710	
35	GGA	ACT	AGT	AGT	GCT	AGT	GGA	ACT	AGT	GGT	GCG	ACT	GGT	TCA	GAT	GGT	624
	Gly	Thr	Ser	Ser	Ala	Ser	Gly	Thr	Ser	Gly	Ala	Thr	Gly	Ser	Asp	Gly	
					715					720					725		
	CAA	ACT	TAC	TCC	ACA	CAA	GCT	ATC	CAA	TAC	CTT	CAA	GGC	CAA	CAA	AAT	672
	Gln	Thr	Tyr	Ser	Thr	Gln	Ala	Ile	Gln	Tyr	Leu	Gln	Gly	Gln	Gln	Asn	
				730				735					740				
40	ATC	TTA	AAT	AAC	GCA	GCG	AAC	TTG	CTC	AAG	CAA	GAT	GAA	TTG	CTC	TTA	720
	Ile	Leu	Asn	Asn	Ala	Ala	Asn	Leu	Leu	Lys	Gln	Asp	Glu	Leu	Leu	Leu	

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	GTT ATT AGC GCT GGG ATA AAC TCC AAC CAA GCT AAC GCT GTG CAA GGG	912
	Val Ile Ser Ala Gly Ile Asn Ser Asn Gln Ala Asn Ala Val Gln Gly	
	810 815 820	
5	CGC GCT AGT CAG CTC CCT AAC GCT CTT TAT AAC GCG CAA GTA ACT TTG	960
	Arg Ala Ser Gln Leu Pro Asn Ala Leu Tyr Asn Ala Gln Val Thr Leu	
	825 830 835	
	GAT AAA ATC AAT GCG CTC AAT AAT CAA GTG AGA AGC ATG CCT TAC TTG	1008
10	Asp Lys Ile Asn Ala Leu Asn Asn Gln Val Arg Ser Met Pro Tyr Leu	
	840 845 850	
	CCC CAA TTC AGA GCC GGG AAC AGC CGT TCA ACG AAT ATT TTA AAC GGG	1056
	Pro Gln Phe Arg Ala Gly Asn Ser Arg Ser Thr Asn Ile Leu Asn Gly	
	855 860 865 870	
15	TTT TAC ACC AAA ATA GGC TAT AAG CAA TTC TTC GGG AAG AAA AGG AAT	1104
	Phe Tyr Thr Lys Ile Gly Tyr Lys Gln Phe Phe Gly Lys Lys Arg Asn	
	875 880 885	
	ATC GGT TTG CGC TAT TAT GGT TTC TTT TCT TAT AAC GGA GCG AGC GTG	1152
20	Ile Gly Leu Arg Tyr Tyr Gly Phe Phe Ser Tyr Asn Gly Ala Ser Val	
	890 895 900	
	GGC TTT AGA TCC ACT CAA AAT AAT GTA GGG TTA TAC ACT TAT GGG GTG	1200
	Gly Phe Arg Ser Thr Gln Asn Asn Val Gly Leu Tyr Thr Tyr Gly Val	
	905 910 915	
25	GGG ACT GAT GTG TTG TAT AAC ATC TTT AGC CGC TCC TAT CAA AAC CGC	1248
	Gly Thr Asp Val Leu Tyr Asn Ile Phe Ser Arg Ser Tyr Gln Asn Arg	
	920 925 930	
	TCT GTG GAT ATG GGC TTT TTT AGC GGT ATC CAA TTA GCC GGT GAG ACC	1296
30	Ser Val Asp Met Gly Phe Phe Ser Gly Ile Gln Leu Ala Gly Glu Thr	
	935 940 945 950	
	TTC CAA TCC ACG CTC AGA GAT GAC CCC AAT GTG AAA TTG CAT GGG AAA	1344
	Phe Gln Ser Thr Leu Arg Asp Asp Pro Asn Val Lys Leu His Gly Lys	
	955 960 965	
35	ATC AAT AAC ACG CAC TTC CAG TTC CTC TTT GAC TTC GGT ATG AGG ATG	1392
	Ile Asn Asn Thr His Phe Gln Phe Leu Phe Asp Phe Gly Met Arg Met	
	970 975 980	
	AAC TTC GGT AAG TTG GAC GGG AAA TCC AAC CGC CAC AAC CAG CAC ACG	1440
40	Asn Phe Gly Lys Leu Asp Gly Lys Ser Asn Arg His Asn Gln His Thr	
	985 990 995	
	GTG GAA TTT GGC GTA GTG GTG CCT ACG ATT TAT AAC ACT TAT TAC AAA	1488
	Val Glu Phe Gly Val Val Val Pro Thr Ile Tyr Asn Thr Tyr Tyr Lys	
	1000 1005 1010	
45	TCA GCA GGG ACT ACC GTG AAG TAT TTC CGT CCT TAT AGC GTT TAT TGG	1536
	Ser Ala Gly Thr Thr Val Lys Tyr Phe Arg Pro Tyr Ser Val Tyr Trp	
	1015 1020 1025 1030	
	TCT TAT GGG TAT TCA TTC TAA	1557
	Ser Tyr Gly Tyr Ser Phe	
	1035	

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 518 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met	Ile	Lys	Lys	Asn	Arg	Thr	Leu	Phe	Leu	Ser	Leu	Ala	Leu	Cys	Ala	1	5	10	15
Ser	Ile	Ser	Tyr	Ala	Glu	Asp	Asp	Gly	Gly	Phe	Phe	Thr	Val	Gly	Tyr	20	25	30	
Gln	Leu	Gly	Gln	Val	Met	Gln	Asp	Val	Gln	Asn	Pro	Gly	Gly	Ala	Lys	35	40	45	
Ser	Asp	Glu	Leu	Ala	Arg	Glu	Leu	Asn	Ala	Asp	Val	Thr	Asn	Asn	Ile	50	55	60	
Leu	Asn	Asn	Asn	Thr	Gly	Gly	Asn	Ile	Ala	Gly	Ala	Leu	Ser	Asn	Ala	65	70	75	80
Phe	Ser	Gln	Tyr	Leu	Tyr	Ser	Leu	Leu	Gly	Ala	Tyr	Pro	Thr	Lys	Leu	85	90	95	
Asn	Gly	Ser	Asp	Val	Ser	Ala	Asn	Ala	Leu	Leu	Ser	Gly	Ala	Val	Gly	100	105	110	
Ser	Gly	Thr	Cys	Ala	Ala	Ala	Gly	Thr	Ala	Gly	Gly	Thr	Ser	Leu	Asn	115	120	125	
Thr	Gln	Ser	Thr	Cys	Thr	Val	Ala	Gly	Tyr	Tyr	Trp	Leu	Pro	Ser	Leu	130	135	140	
Thr	Asp	Arg	Ile	Leu	Ser	Thr	Ile	Gly	Ser	Gln	Thr	Asn	Tyr	Gly	Thr	145	150	155	160
Asn	Thr	Asn	Phe	Pro	Asn	Met	Gln	Gln	Gln	Leu	Thr	Tyr	Leu	Asn	Ala	165	170	175	
Gly	Asn	Val	Phe	Phe	Asn	Ala	Met	Asn	Lys	Ala	Leu	Glu	Asn	Lys	Asn	180	185	190	
Gly	Thr	Ser	Ser	Ala	Ser	Gly	Thr	Ser	Gly	Ala	Thr	Gly	Ser	Asp	Gly	195	200	205	
Gln	Thr	Tyr	Ser	Thr	Gln	Ala	Ile	Gln	Tyr	Leu	Gln	Gly	Gln	Gln	Asn	210	215	220	
Ile	Leu	Asn	Asn	Ala	Ala	Asn	Leu	Leu	Lys	Gln	Asp	Glu	Leu	Leu	Leu	225	230	235	240
Glu	Ala	Phe	Asn	Ser	Ala	Val	Ala	Ala	Asn	Ile	Gly	Asn	Lys	Glu	Phe	245	250	255	
Asn	Ser	Ala	Ala	Phe	Thr	Gly	Leu	Val	Gln	Gly	Ile	Ile	Asp	Gln	Ser	260	265	270	
Gln	Ala	Val	Tyr	Asn	Glu	Leu	Thr	Lys	Asn	Thr	Ile	Ser	Gly	Ser	Ala	275	280	285	
Val	Ile	Ser	Ala	Gly	Ile	Asn	Ser	Asn	Gln	Ala	Asn	Ala	Val	Gln	Gly	290	295	300	

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Arg Ala Ser Gln Leu Pro Asn Ala Leu Tyr Asn Ala Gln Val Thr Leu
 305 310 315 320
 5 Asp Lys Ile Asn Ala Leu Asn Asn Gln Val Arg Ser Met Pro Tyr Leu
 325 330 335
 Pro Gln Phe Arg Ala Gly Asn Ser Arg Ser Thr Asn Ile Leu Asn Gly
 340 345 350
 10 Phe Tyr Thr Lys Ile Gly Tyr Lys Gln Phe Phe Gly Lys Lys Arg Asn
 355 360 365
 Ile Gly Leu Arg Tyr Tyr Gly Phe Phe Ser Tyr Asn Gly Ala Ser Val
 370 375 380
 15 Gly Phe Arg Ser Thr Gln Asn Asn Val Gly Leu Tyr Thr Tyr Gly Val
 385 390 395 400
 Gly Thr Asp Val Leu Tyr Asn Ile Phe Ser Arg Ser Tyr Gln Asn Arg
 405 410 415
 20 Ser Val Asp Met Gly Phe Phe Ser Gly Ile Gln Leu Ala Gly Glu Thr
 420 425 430
 Phe Gln Ser Thr Leu Arg Asp Asp Pro Asn Val Lys Leu His Gly Lys
 435 440 445
 25 Ile Asn Asn Thr His Phe Gln Phe Leu Phe Asp Phe Gly Met Arg Met
 450 455 460
 Asn Phe Gly Lys Leu Asp Gly Lys Ser Asn Arg His Asn Gln His Thr
 465 470 475 480
 30 Val Glu Phe Gly Val Val Val Pro Thr Ile Tyr Asn Thr Tyr Tyr Lys
 485 490 495
 Ser Ala Gly Thr Thr Val Lys Tyr Phe Arg Pro Tyr Ser Val Tyr Trp
 500 505 510
 35 Ser Tyr Gly Tyr Ser Phe
 515

40 Claims

1. A recombinant attenuated microbial pathogen, which comprises at least one heterologous nucleic acid molecule encoding a Helicobacter antigen, wherein said pathogen is capable to express said nucleic acid molecule or capable to cause the expression of said nucleic acid molecule in a target cell.
2. The pathogen according to claim 1, which is an enterobacterial cell, especially a Salmonella cell.
3. The pathogen according to claim 1 or 2, which is a Salmonella aro mutant cell.
4. The pathogen according to any of claims 1-3, wherein the Helicobacter antigen is urease, a urease subunit, an immunologically reactive fragment thereof, or a peptide mimotope thereof.
5. The pathogen according to any one of claims 1-3, wherein the Helicobacter antigen is a secretory polypeptide from Helicobacter, an immunologically reactive fragment thereof, or a peptide mimotope thereof.
6. The pathogen according to any one of claims 1-3 and 5, wherein the Helicobacter antigen is selected from the

group consisting of the antigens AlpA, AlpB, immunologically reactive fragments thereof, or a peptide mimotope thereof.

- 5 7. The pathogen according to any one of claims 1-6, wherein said nucleic acid molecule encoding a Helicobacter antigen is capable to be expressed phase variably.
8. The pathogen according to claim 7,
wherein said nucleic acid molecule encoding the Helicobacter antigen is under control of an expression signal which is substantially inactive in the pathogen and which is capable to be activated by a nucleic acid reorganization
10 caused by a nucleic acid reorganization mechanism in the pathogen.
9. The pathogen according to claim 8,
wherein the expression signal is a bacteriophage promoter, and the activation is caused by a DNA reorganization
15 resulting in the production of a corresponding bacteriophage RNA polymerase in the pathogen.
10. The pathogen according to any one of claims 1-9, further comprising at least one second nucleic acid molecule
encoding an immunomodulatory polypeptide, wherein said pathogen is capable to express said second nucleic
acid molecule.
- 20 11. Pharmaceutical composition comprising as an active agent a recombinant attenuated pathogen according to any
one of claims 1-10, optionally together with pharmaceutically acceptable diluents, carriers and adjuvants.
12. Composition according to claim 11, which is a living vaccine, which is suitable for administration to a mucosal sur-
face or via the parenteral route.
- 25 13. A method for the preparation of a living vaccine comprising formulating an attenuated pathogen according to any
one of claims 1-10 in a pharmaceutically effective amount with pharmaceutically acceptable diluents, carriers
and/or adjuvants.
- 30 14. A method for preparing a recombinant attenuated pathogen according to any one of claims 1-10, comprising the
steps:
 - a) inserting a nucleic acid molecule encoding a Helicobacter antigen into an attenuated pathogen, wherein a
35 recombinant attenuated pathogen is obtained, which is capable of expressing said nucleic acid molecule or is
capable to cause expression of said nucleic acid molecule in a target cell, and
 - b) cultivating said recombinant attenuated pathogen under suitable conditions.
15. The method according to claim 15,
40 wherein said nucleic acid molecule encoding a Helicobacter antigen is located on an extrachromosomal plasmid.
16. A method for identifying Helicobacter antigens, which raise a protective immune response in a mammalian host,
comprising the steps of:
 - 45 a) providing an expression gene bank of Helicobacter in an attenuated pathogen and
 - b) screening the clones of the gene bank for their ability to confer protective immunity against a Helicobacter
infection in a mammalian host.

FIG. 1

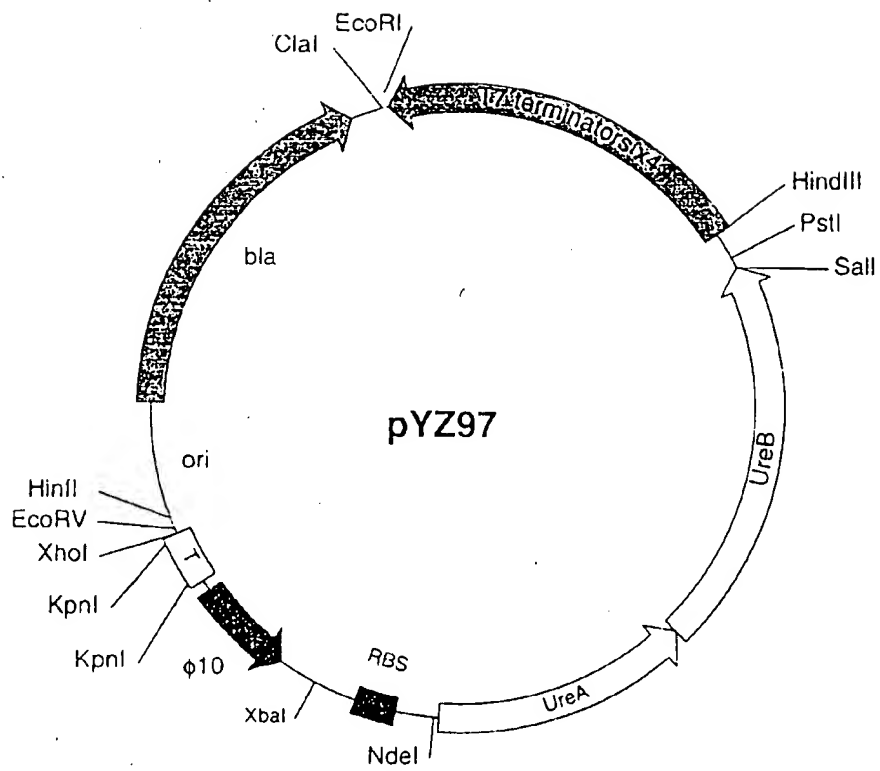
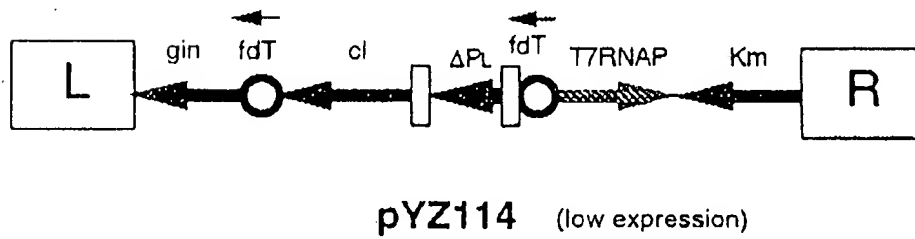
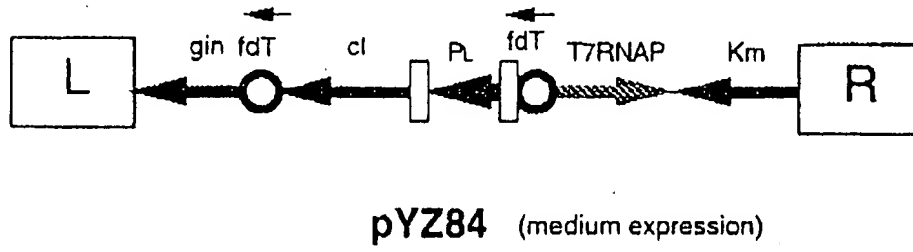
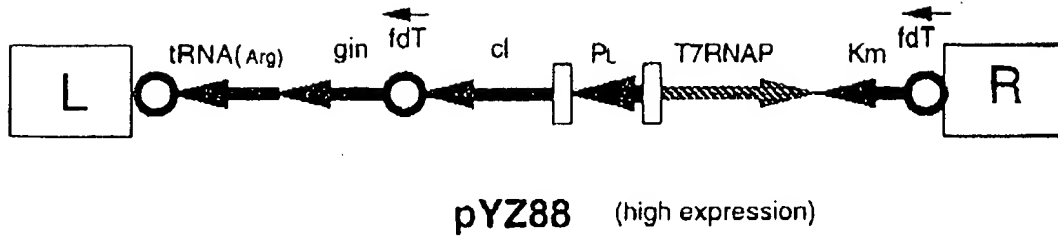


FIG. 2



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 96 11 6337

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
T	WO 96 33732 A (ORAVAX INC) 31 October 1996 * the whole document *		C12N1/21 C12N15/31 C12N15/74 A61K39/02 A61K39/112 C12Q1/68
A	EP 0 654 273 A (LEVEEN HARRY H ; LAVEEN ERIC G (US); LAVEEN ROBERT F (US)) 24 May 1995 * column 11, line 26 - column 12, line 40; claim 41 *	1-16	
A	INFECT. IMMUN. (1994), 62(11), 4981-9 CODEN: INFIBR; ISSN: 0019-9567, 1994, XP002011225 FERRERO, RICHARD L. ET AL: "Recombinant antigens prepared from the urease subunits of Helicobacter spp.: evidence of protection in a mouse model of gastric infection" * page 4984, Expression of Helicobacter urease polypeptides in E. coli *	1-16	
			TECHNICAL FIELDS SEARCHED (Int.Cl.6)
			C12N A61K C12Q
The present search report has been drawn up for all claims			
Place of search MUNICH		Date of completion of the search 8 April 1997	Examiner Halle, F
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application I : document cited for other reasons A : member of the same patent family, corresponding document</p>			